

TETRAHEDRON LETTERS

Tetrahedron Letters 44 (2003) 7091-7094

## A new efficient synthetic process for the construction of the pentacyclic core of marine alkaloid ecteinascidins

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Received 13 May 2003; revised 27 June 2003; accepted 14 July 2003

Abstract—The pentacyclic core of ecteinascidins was constructed from two fundamental building blocks, the 1,2,3,4-tetra-hydroisoquinoline derivative and the substituted phenylalanine derivative, via 8 steps using readily available L-Dopa as starting material. The key steps involve coupling of the two aforementioned building blocks, regioselective reduction of the 11-carbonyl group of the key intermediate piperazine-1,4-dione derivative, and intramolecular Pictet-Spengler cyclization.

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Ecteinascidins are a family of marine-derived tetrahydroisoquinoline alkaloids, which possess extremely potent antitumor activity. Ecteinascidin 743 (Et 743, 1), the most abundant and bioactive of this family, is being studied in phase I/II clinical trials. Much synthetic effort has been directed towards the synthesis of Et 743, but due to its formidable structural complexity, there have been only three successful results reported so far by Corey, Cuevas and Fukuyama groups, respectively. Fortunately, phthalascidin (Pt 650, 2), a structurally simplified version of 1, was found to exhibit comparable antitumor activity to that of Et 743 and may therefore be a more practical therapeutic agent. In

view of the structural feature, the cores of both 1 and 2 are characterized by a bis(tetrahydroisoquinoline) pentacyclic system (ABCDE), which is believed to be the primary pharmacophore through the preliminary structure–activity study of Et 743 and its analogs (Fig. 1).<sup>5</sup> This inspired us to start a research project on the discovery of structurally simplified, synthetically accessible and biologically active ecteinascidin analogs as antitumor agents. In this communication, we report the synthesis of 3 that bears the same pentacyclic framework as that of 1 and 2 via a new efficient synthetic route, which we believe is of considerable value in the synthesis of simplified ecteinascidin analogs.

Figure 1.

Keywords: ecteinascidins; pentacyclic core; L-Dopa; Pictet-Spengler cyclization.

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Retrosynthetic analysis of the pentacyclic core of ecteinascidins leads us to employ an original synthetic strategy different from those reported.<sup>3</sup> According to our design, the BCD-ring system could be constructed sequentially (Scheme 1). Thus, 4 could be derived from 5 through reduction of the 21-carbonyl group followed by replacement of the hydroxyl group with the cyanide group. Compound 5 could be obtained from 6 via an intramolecular Pictet-Spengler cyclization, and 6 maybe accessed by regioselective reduction of the 11carbonyl group of the piperazine-1,4-dione derivative 7, a key intermediate in our synthesis. Compound 7, as we assumed, could be derived from two fundamental building blocks, the 1,2,3,4-tetrahydroisoquinoline derivative 8 and the substituted phenylalanine derivative 9, via formation of two amide bonds. Both 8 and 9 could be easily obtained from the substituted phenylalanine 10 by several functional group manipulations.

Partly enlightened by the biosynthetic study on ecteinascitins,<sup>6</sup> which supposed that the pentacyclic core of ecteinascidins was derived from two molecules of L-Dopa or L-tyrosine, we chose the commercially available L-Dopa as the starting material. Thus, protection of the amino group of L-Dopa by BOC provided 11, which was treated with an excess of Me<sub>2</sub>SO<sub>4</sub>/K<sub>2</sub>CO<sub>3</sub> in acetone at reflux to give 12.<sup>7</sup> Then hydrolysis of 12 in 1N NaOH afforded 13 as one of the two building blocks. The overall yield of 13 from L-Dopa was 70% (Scheme 2).

According to a known method, L-Dopa was submitted to Pictet-Spengler reaction using 37% HCHO in

0.5N H<sub>2</sub>SO<sub>4</sub> to yield the 6,7-dihydroxy-1,2,3,4-tetrahydroisoguinoline-3-carboxylic acid 14. Of particular note, formaldehyde was chosen here for convenience. We have investigated a series of aliphatic and aromatic aldehydes in this reaction to produce the corresponding 1-substituded tetrahydroisoguinoline derivatives. In each case, the 1,3-cis isomer was obtained as the major product.9 Thus, it is feasible to introduce a desired stereocenter on the 1-postion of the pentacyclic core via an asymmetric Pictet-Spengler reaction when needed. Esterification of 14 in HCl/CH<sub>3</sub>OH at reflux provided 15. Protection of the nitrogen of 15 by a formyl group afforded 16, which was transformed into the methylation product 17 upon treatment with Me<sub>2</sub>SO<sub>4</sub>/K<sub>2</sub>CO<sub>3</sub> in acetone at reflux. Cleavage of the formyl group in HCl/CH<sub>3</sub>OH provided another building block 18.<sup>10</sup> The overall yield of 18 from L-Dopa is ca. 50% (Scheme 3).

Coupling of 13 and 18 to form 19 was accomplished through the action of bis(2-oxo-3-oxazolidinyl) phosphinic chloride (BOP-Cl) in 80% yield. Then 19 was treated with CF<sub>3</sub>CO<sub>2</sub>H in CH<sub>2</sub>Cl<sub>2</sub> to afford the key intermediate piperazine-1,4-dione derivative 20, which was converted to the carbamate 21 in 80% yield according to a known procedure. Regioselective reduction of 11-carbonyl group of 21 with an excess amount of Li(*t*-BuO)<sub>3</sub>AlH in THF provided the diastereomeric mixture of the alcohol 22, which, on treatment with HCO<sub>2</sub>H at 70°C, yielded exclusively the expected pentacyclic framework of ecteinascidins 23. It is worth noting that a compound similar to 23 was committed to the same cyclization condition by another research team,

$$\begin{array}{c} AB,C \\ AB$$

Scheme 1.

Scheme 2. Reagents and conditions: (a) (Boc)<sub>2</sub>O, Et<sub>3</sub>N, DMF, 60°C, 89%; (b) Me<sub>2</sub>SO<sub>4</sub>, K<sub>2</sub>CO<sub>3</sub>, CH<sub>3</sub>COCH<sub>3</sub>, reflux, 92%; (c) 1N KOH/CH<sub>3</sub>OH, 90%.

HO 
$$NH_2$$
  $NH_2$   $NH_2$   $NH_3$   $NH_4$   $NH_4$   $NH_4$   $NH_5$   $NH_6$   $NH_6$ 

**Scheme 3.** Reagents and conditions: (a) HCHO, 0.5N H<sub>2</sub>SO<sub>4</sub>, 24 h, 75%; (b) HCl/CH<sub>3</sub>OH, reflux, 6 h, 99%; (c) Ac<sub>2</sub>O/HCO<sub>2</sub>H, 4 h; then H<sub>2</sub>O/CH<sub>3</sub>OH, 87%; (d) Me<sub>2</sub>SO<sub>4</sub>, K<sub>2</sub>CO<sub>3</sub>, CH<sub>3</sub>COCH<sub>3</sub>, reflux, 90%; (e) HCl/CH<sub>3</sub>OH, reflux, 4 h, 85%.

$$\begin{array}{c} \text{CH}_{3O} \\ \text{CH}_{3O} \\ \text{CH}_{3O} \\ \text{20} \end{array} \\ \begin{array}{c} \text{CH}_{3O} \\ \text{CH}_{3O} \\ \text{CH}_{3O} \\ \text{OCH}_{3} \end{array} \\ \begin{array}{c} \text{CH}_{3O} \\ \text{CH}_{3O} \\ \text{OCH}_{3} \end{array} \\ \begin{array}{c} \text{CH}_{3O} \\ \text{OCH}_{3} \\ \text{OCH}_{3} \\ \text{OCH}_{3} \end{array} \\ \begin{array}{c} \text{CH}_{3O} \\ \text{OCH}_{3} \\ \text{OCH}_{4} \\ \text{OCH}_{4} \\ \text{OCH}_{5} \\$$

$$\begin{array}{c} OCH_3 \\ OCH_3 \\$$

Scheme 4. Reagents and conditions: (a) BOP-Cl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 4 h, 80%; (b) CF<sub>3</sub>COOH/CH<sub>2</sub>Cl<sub>2</sub>, 2 h, 86%; (c) ClCO<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>, DMAP, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 6 h, 80%; (d) Li(t-BuO)<sub>3</sub>AlH, THF, 0°C, 2 h, then H<sub>2</sub>O, 90%; (e) HCO<sub>2</sub>H, 70°C, 3 h, 65%; (f) H<sub>2</sub>SO<sub>4</sub>/CF<sub>3</sub>CO<sub>2</sub>H, 15 h, 98%; (g) HCHO/HCO<sub>2</sub>H, 70°C, 2 h, 95%; (h) LiAlH<sub>4</sub>, THF, -17°C, 0.5 h, then 0°C, 1 h; KCN, phosphate buffer (pH=7), 2 h, 82%.

but an unsuccessful result was reported.<sup>12</sup> Next, deprotection of **23** with a mixture of CF<sub>3</sub>CO<sub>2</sub>H and H<sub>2</sub>SO<sub>4</sub> gave the secondary amine **24** in quantitative yield. Reductive methylation of **24** with HCHO/HCO<sub>2</sub>H at 70°C for 2 h provided **25** in 95% yield. Finally, the lactam ring of **25** could be easily reduced through treatment with an excess of LiAlH<sub>4</sub> in THF at 0°C for 1 h to afford the corresponding cyclic hemiaminal, which upon exposure to KCN in phosphate buffer (pH=7) afforded the pentacyclic amino nitrile **3** as an enantiomerically pure product in 82% overall yield from **25** (Scheme 4).

Gratifyingly, 3 does not only possess the desired pentacyclic core of ecteinascidins, but also bears four completely correct stereocenters (3S,11S,13S,21R) as those of the natural products. This conclusion was verified on the basis of its spectroscopic data, <sup>13</sup> especially the NOE difference spectroscopy. Obvious NOE enhancement

was observed between H-3 and H-11, thus the *syn* C3–C11 backbone stereochemical relationship was established. Similarly, irradiation of the H-21 produced noticeable NOE enhancement of H $\beta$ -1,but not of H $\alpha$ -1,which indicated that H-21 was of  $\beta$ -configuration

$$\begin{array}{c} \text{CH}_3\text{O} \\ \text{OCH}_3 \\ \text{OCH}_3 \\ \text{OCH}_3 \\ \text{H}_{\beta} \\ \text{H}_{\alpha} \\ \text{NOE} \\ \text{NOE} \\ \text{NOE} \\ \text{OCH}_3 \\ \text{OCH}_$$

Figure 2.

(Fig. 2). These data are consistent with the stereochemical relationship of the four stereocenters.

In summary, we have developed a new efficient synthetic route for the construction of the pentacyclic core of ecteinascidins starting from L-Dopa. Further study on the synthesis of structurally simplified and bioactive ecteinascidin analogs based on this methodology is ongoing in our laboratory.

## References

- For a review, see: Scott, J. D.; Williams, R. M. Chem. Rev. 2002, 102, 1669–1730.
- 2. For clinical data, see: (a) Ryan, D. P.; Supko, J. G.; Eder, J. P.; Seiden, M. V.; Demetri, G.; Lynch, T. J.; Fischman, A. J.; Davis, J.; Jimeno, J.; Clark, J. W. Clin. Cancer Res. 2001, 7, 231-242; (b) Zelek, L.; Yovine, A.; Etienne, B.; Jimeno, J.; Taamma, A.; Martin, C.; Spielmann, M.; Cvitkovic, E.; Misset, J. L. The American Society of Clinical Oncology, 36th Annual Meeting, New Orleans, May 20-23, 2000; Abstract number 592; (c) Delaloge, S.; Yovine, A.; Taamma, A.; Cottu, P.; Riofrio, M.; Raymond, E.; Brain, E.; Marty, M.; Jimeno, J.; Cvitkovic, E.; Misset, J. L. The American Society of Clinical Oncology, 36th Annual Meeting, New Orleans, May 20-23, 2000; Abstract number 2181; (d) Le Cesne, A.; Judson, I.; Blay, J. Y.; Radford, J.; van Oosterom, A.; Lorigan, P.; Rodenhuis, E.; Donato Di Paoula, E.; Van Glabbeke, M.; Jimeno, J.; Verweij, J. The American Society of Clinical Oncology, 36th Annual Meeting, New Orleans, May 20-23, 2000; Abstract number 2182; (e) Aune G. J.; Furuta T.; Pommier Y. Anti-cancer Drugs 2002, 13, 545-555.
- For some recent examples, see: (a) Saito, N.; Tachi, M.; Seki, R.; Kamayachi, H.; Kubo, A. Chem. Pharm. Bull. 2000, 48,1549–1557; (b) Endo, A.; Kann, T.; Fukuyama, T. Synlett 1999, 1103–1105; (c) Zhou, B.; Guo J.; Danishefsky, S. J. Tetrahedron Lett. 2000, 41, 2043–2046; (d) Martinez, E. J.; Corey, E. J. Org. Lett. 2000, 2, 993–996; (e) Jin, W.; Metobo, S.; Williams, R. M. Org. Lett. 2003, in press.

- (a) Corey, E. J.; Gin, D. Y.; Kania, R. S. J. Am. Chem. Soc. 1996, 118, 9202–9203; (b) Cuevas, C.; Pérez, M.; Martín, M. J.; Chicharro, J. L.; Fernádez-Rivas, C.; Flores, M.; Francesch, A.; Gallego, P.; Zarzuelo, M.; de la Calle, F.; García, J.; Polanco, C.; Rodríguez, I.; Manzanares, I. Org. Lett. 2000, 2, 2545–2548; (c) Endo, A.; Yanagisawa, A.; Abe, M.; Tohma, S.; Kan, T.; Fukuyama, T. J. Am. Chem. Soc. 2002, 124, 6552–6554.
- Martinez, E. J.; Owa, T.; Schreiber, S. L.; Corey, E. J. Proc. Natl. Acad. Sci. 1999, 96, 3496–3501.
- (a) Kerr, R. G.; Miranda, N. F. J. Nat. Prod. 1995, 58, 1618–1621;
   (b) Sakai, R.; Jares-Erijman, E. A.; Manzanares, I.; Elipe, M. V. S.; Rinehart, K. L. J. Am. Chem. Soc. 1996, 118, 9017–9023.
- 7. Garciá, E.; Arrasate, S.; Ardeo, A.; Lete, E.; Sotomayor, N. *Tetrahedron Lett.* **2001**, *42*, 1511–1513.
- 8. Brossi, A.; Focella, A.; Teitel, S. Helv. Chim. Acta 1972, 55, 15-21.
- 9. Wang, Y.; Liu, Z. Z.; Chen, S. Z.; Liang, X. T. Chin. Chem. Lett. 2003, in press.
- Dean, R. T.; Rapoport, H. J. Org. Chem. 1978, 43, 4183–4189.
- Kubo, A.; Saito, N.; Yamato, H.; Masubuchi, K.; Nakamura, M. J. Org. Chem. 1988, 53, 4295–4310.
- Fukuyama, T.; Sacheben, R. A. J. Am. Chem. Soc. 1982, 104, 4957–4958.
- 13. Analytical and spectroscopic data for 3: light yellow solid; mp 124.5–127°C;  $[\alpha]_D^{20} = +19.3$  (c 0.4, CHCl<sub>3</sub>); <sup>1</sup>H NMR (300 Hz, CDCl<sub>3</sub>):  $\delta$  2.35 (s, 3H, -NCH<sub>3</sub>), 2.64–2.55 (m, 3H, H-4+H-14), 3.14 (dd, 1H, J=8.4, 18.0 Hz, H-4), 3.24-3.18 (ddd, 1H, J=3.0, 4.8, 8.4 Hz, H-3), 3.47 (d, 1H, J=8.1 Hz, H-13), 3.59 (d, 1H, J=3.0 Hz, H-11), 3.71 (d, 1H, J=15.0 Hz, H $\beta$ -1), 3.78–3.81 (m, 9H, 3× CH<sub>3</sub>O), 3.87 (d, 1H, J=15.0Hz, Ha-1), 3.90 (s, 3H,  $CH_3O$ ), 3.94 (d, 1H, J=2.1 Hz, H-21), 6.43 (s, 1H, Ar-H), 6.48 (s, 1H, Ar-H), 6.52 (s, 1H, Ar-H), 6.57 (s, 1H, Ar-H);  ${}^{13}$ C NMR (300 Hz, CDCl<sub>3</sub>):  $\delta$  25.7, 31.6, 42.0, 54.2, 55.4, 55.8, 55.9, 55.9, 56.3, 56.3, 62.2, 62.4, 108.6, 110.8, 110.9, 113.5, 117.2, 123.5, 123.8, 124.3, 126.1, 146.5, 147.4, 147.7, 148.4; IR (KBr, cm<sup>-1</sup>): 2220 (CN, w), 1660 (C=O, s); FAB-MS (m/z): 436 (M+1, 27%), 435 (M, 10%), 409 (22%), 204 (100%); HRMS (FAB): calcd for C<sub>25</sub>H<sub>30</sub>N<sub>3</sub>O<sub>4</sub> (M+1) 436.2236; found 435.2214.